## Screening for NERDs in *Arabidopsis thaliana*

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#### **Abstract:**

Plant survival depends upon the directed growth of plant roots into the soil, where essential water and nutrients can be obtained. The molecular basis of this root growth depends upon a protein complex called the exocvst. Mutations affecting exocvst proteins result in slow root growth and dwarf roots (see figure to right). In non-plant species the exocyst has been shown to be involved in exocytosis and secretion, but exactly how the exocyst functions to affect root growth is unknown. To better understand the exocyst's role in root growth, a screen was developed to identify proteins that interact with the exocyst. These interacting proteins are named New Enhancers of Root Dwarfism, or NERDs. I worked on the last step of the screen for NERDs, searching the phenotypes and genotypes of over 6,000 Arabidopsis plants in 21 families for signs of an interaction. Two families yielded interesting results, potentially identifying two new NERDs involved with the exocyst in determining the rate and direction of root growth, as well as the formation and morphology of root hairs. My results indicate the genetics involved in identifying these NERDs is more complicated than anticipated, and provide a direction for future investigations.



mutation (sec8-3) affects root development

#### The Nerd Screen:

The goal of the NERD Screen is the identification of novel exocyst interactors. Through EMS exposure. random mystery mutations were introduced into many lines homozygous for the sec8-6 mutation. These lines were manipulated to isolate those new mutations that interacted synergistically with sec8-6. The last step of the screen, and the step that I carried out this summer, is called M5 verification. If an exocyst interactor was mutated through EMS exposure, a severe mutant phenotype would occur in one in sixteen generation M5 plants, which would be homozygous for the mystery mutation and sec8-6.

	sec8-6	mystery mutatior		
мо	ee	++		
	↓ EMS Mu	tagenesis $\downarrow$		
M1	ee	a+		
	$\downarrow$ Self Cross $\downarrow$			
M2	ee	aa, a+, ++		
	↓Phenotypi	c Selection↓		
M3	ee	aa		
	↓ Outcross to	o Wild Type↓		
M4	e+	a+		
	↓ Self	Cross ↓		
M5	ee,e+,++	aa, a+, ++		

Verification



e: sec8-6 (exocyst mutation) allele, a: mutant allele from EMS mutagenesis, +: wild type allele

## Family 9 Phenotype S:

Plants with phenotype S do not have any root hairs, and have squiggly roots. Plants with this phenotype are 90% homozygous for sec8-6 This is significantly greater than the 25% expected for a non interactor. Alternative hypotheses ar discussed below. Phenotype S Observed: 56/332 or ~2.7/16

Phenotype S	Wild Ty	Vild Type					Phenotype S
		Family 9 Genotypes					
		ee		e+		++	
Phenotype	Total	#	%	#	%	#	%
Negative Result			25%		50%		25%
WT	9	1	11%	5	56%	3	33%
C C	20	18	QU0%	2	10%	Λ	<u>∩%</u>

Total

13 1

15 13

Plants with phenotype G have shorter roots with short, dense, abnormally shaped root hairs. Phenotype G plants are 87% homozygous for sec8-6. This is significantly greater than the 25% expected for a non-interactor. Alternative hypotheses are discussed below. Phonotypo C

Observed		
50/299 or		
~2.7/16		
/.•		

sec8-6 and the mystery mutations that cause phenotypes S and G may be linked genes. This means that they are located near each other on the same chromosome, increasing their chances of being inherited together. Crossing over may allow for some independent assortment. If this hypothesis is confirmed, the mutated proteins that cause these phenotypes may not be exocyst interactors.



# S 2018 90% 2 10% 0 0%

#### Family 10 Phenotype G:

ee

%

25%

8%

87%

#

i nenotype o j	
Observed:	,
50/299 or	
~2.7/16	

### **Hypothesis A:**

Phenotype

legative Result

WΤ

G





Family 10 Genotypes

e+

%

8 62%

2 13%

**Hypothesis B:** 

The mystery mutations that cause phenotypes

S and G may act as dominant alleles, meaning

50%

#

++

%

25%

31%

0%

#

4

0

nutation) allele, **a**: mutant allele from EMS mutagenesis, +: wild type allele

#### **Procedure:**

•Prepared plates with growth medium under a laminar flow hood •Put seeds onto plates with a micropipettor

·Grew plates upright in a climatecontrolled growth chamber Made visual observations with the aid of a dissecting microscope on days three, five, and seven •Performed DNA extraction with the aid of a drill and special bit

 Amplified DNA by PCR on a RoboCvcler

 Imaged DNA following gel electrophoresis

#### **Results:**

Family	Outcome	Family	Outcome
I	appears to be 1/4	12	I/I6 category not valid
2	genotyping negative	13	appears to be 1/4
3	genotyping negative	14	appears to be 1/4
4	too many phenotypes, dead	15	sec8-6 not present
5	genotyping negative	16	sec8-6 not present
6	genotyping negative	17	sec8-6 not present
7	chlorosis	18	sec8-6 not present
8	genotyping negative	19	sec8-6 not present
9	putative NERD	20	sec8-6 not present
10	putative NERD	21	sec8-6 not present
	no 1/16 category		

#### **Conclusion:**

Families 9 and 10 are putative NERDs, however additional research is required to confirm and investigate this finding.

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